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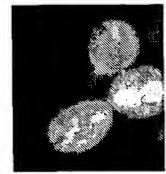
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Antibodies - General Information

This page summarises a lot of generally useful information about antibodies.

The Kabat Numbering Scheme

The Kabat numbering scheme is a widely adopted standard for numbering the residues in an antibody in a consistent manner. However the scheme does have its problems!

First, since the numbering scheme was developed from (fairly limited) sequence data, the position at which insertions occur in CDR-L1 and CDR-H1 does not match the structural insertion position. Thus topologically equivalent residues in these loops do not get the same number.

Second, the numbering adopts a rigid specification. For example in the potentially very long CDR-H3, insertions are numbered between residue H100 and H101 with letters up to K (i.e. H100, H100A ... H100K, H101). If there are more residues than that, there is no standard way of numbering them. Such situations occur at other positions too.

The numbering throughout the chains follows.

Light chain

0	1	2	3	4	5	6	7	8	9
10	11	12	13	14	15	16	17	18	19
20	21	22	23	24	25	26	27		
27A	27B	27C	27D	27E	27F			28	29
30	31	32	33	34	35	36	37	38	39
40	41	42	43	44	45	46	47	48	49
50	51	52	53	54	55	56	57	58	59
60	61	62	63	64	65	66	67	68	69
70	71	72	73	74	75	76	77	78	79
80	81	82	83	84	85	86	87	88	89
90	91	92	93	94	95				
95A	95B	95C	95D	95E	95F	96	97	98	99
100	101	102	103	104	105	106			
106A							107	108	109

Heavy chain

0	1	2	3	4	5	6	7	8	9
10	11	12	13	14	15	16	17	18	19
20	21	22	23	24	25	26	27	28	29
30	31	32	33	34	35				
35A	35B					36	37	38	39
40	41	42	43	44	45	46	47	48	49
50	51	52							
52A	52B	52C	53	54	55	56	57	58	59
60	61	62	63	64	65	66	67	68	69
70	71	72	73	74	75	76	77	78	79
80	81	82							
82A	82B	82C	83	84	85	86	87	88	89
90	91	92	93	94	95	96	97	98	99
100									
100A	100B	100C	100D	100E	100F	100G	100H	100I	100J
100K	101	102	103	104	105	106	107	108	109

110 111 112 113

The Chothia Numbering Scheme

The Chothia numbering scheme is identical to the Kabat scheme, but places the insertions in CDR-L1 and CDR-H1 at the structurally correct positions. This means that topologically equivalent residues in these loops do get the same label (unlike the Kabat scheme). There are two disadvantages: first, the Kabat scheme is so widely used that some confusion can arise; second, Chothia **et al. changed** their numbering scheme as of their 1989 Nature paper such that insertions in CDR-L1 are placed after residue L31 rather than L30. Examining the conformations of the loops shows that **L30 is the correct position**.

The pre-1989 Chothia numbering (the structurally correct version) throughout the chains follows.

Note That in their latest paper (Al-Lazikani et al., (1997) JMB **273**,927-948), Chothia's group returns to using residue 30 as the insertion site in CDR-L1!

Light chain

0	1	2	3	4	5	6	7	8	9
10	11	12	13	14	15	16	17	18	19
20	21	22	23	24	25	26	27	28	29
30									
30A	30B	30C	30D	30E	30F				
	31	32	33	34	35	36	37	38	39
40	41	42	43	44	45	46	47	48	49
50	51	52	53	54	55	56	57	58	59
60	61	62	63	64	65	66	67	68	69
70	71	72	73	74	75	76	77	78	79
80	81	82	83	84	85	86	87	88	89
90	91	92	93	94	95				
95A	95B	95C	95D	95E	95F	96	97	98	99
100	101	102	103	104	105	106			
106A							107	108	109

Heavy chain

0	1	2	3	4	5	6	7	8	9
10	11	12	13	14	15	16	17	18	19
20	21	22	23	24	25	26	27	28	29
30	31								
31A	31B								
		32	33	34	35	36	37	38	39
40	41	42	43	44	45	46	47	48	49
50	51	52							
52A	52B	52C	53	54	55	56	57	58	59
60	61	62	63	64	65	66	67	68	69
70	71	72	73	74	75	76	77	78	79
80	81	82							
82A	82B	82C	83	84	85	86	87	88	89
90	91	92	93	94	95	96	97	98	99
100									
100A	100B	100C	100D	100E	100F	100G	100H	100I	100J
100K	101	102	103	104	105	106	107	108	109
110	111	112	113						

Table of CDR Definitions

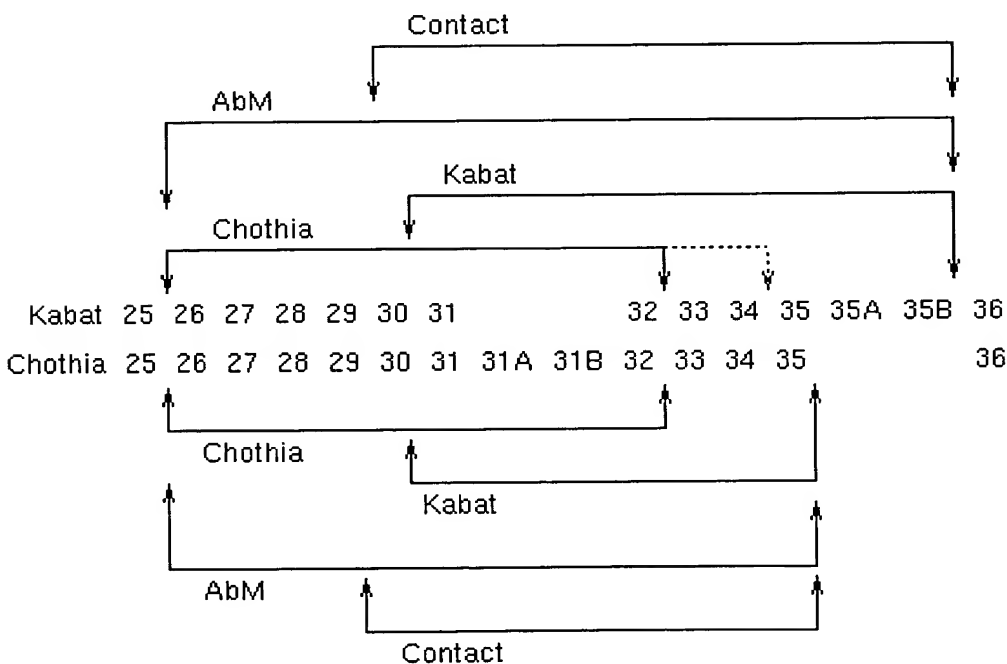
A number of definitions of the CDRs are commonly in use:

- The Kabat definition is based on sequence variability and is the most commonly used.
- The Chothia definition is based on the location of the structural loop regions.
- The AbM definition is a compromise between the two used by Oxford Molecular's AbM antibody modelling software.
- The contact definition has been recently introduced by us and is based on an analysis of the available complex crystal structures. This definition is likely to be the most useful for people wishing to perform mutagenesis to modify the affinity of an antibody since these are residues which take part in interactions with antigen.

Loop	Kabat	AbM	Chothia	Contact
L1	L24 -- L34	L24 -- L34	L24 -- L34	L30 -- L36
L2	L50 -- L56	L50 -- L56	L50 -- L56	L46 -- L55
L3	L89 -- L97	L89 -- L97	L89 -- L97	L89 -- L96
H1	H31 -- H35B (Kabat Numbering)	H26 -- H35B	H26 -- H32..34	H30 -- H35B
H1	H31 -- H35 (Chothia Numbering)	H26 -- H35	H26 -- H32	H30 -- H35
H2	H50 -- H65	H50 -- H58	H52 -- H56	H47 -- H58
H3	H95 -- H102	H95 -- H102	H95 -- H102	H93 -- H101

Note that the end of the Chothia CDR-H1 loop when numbered using the Kabat numbering convention varies between H32 and H34 depending on the length of the loop. (This is because the Kabat numbering scheme places the insertions at H35A and H35B.)

- If neither 35A nor 35B is present, the loop ends at 32
- If only 35A is present, the loop ends at 33
- If both 35A and 35B are present, the loop ends at 34



This diagram illustrates the alternative definitions for CDR-H1. The Kabat and Chothia numbering schemes are shown horizontally and the Kabat, Chothia, AbM and Contact definitions of the CDRs are shown with arrows above and below the two numbering schemes.

Table of mean contact data

Following an analysis of the contacts between antibody and antigen in the complex structures available in the Protein Databank, we have generated a set of mean contact data. The full method by which these results were obtained is described in the following paper: MacCallum, R. M., Martin, A. C. R. and Thornton, J. T. **Antibody-antigen interactions: Contact analysis and binding site topography.** J. Mol. Biol. **262**, 732-745.

Briefly, we have analysed the number of contacts made at each position, defining contact as burial by > 1 square Angstrom change in solvent accessibility. These data give a simple measure of how likely a residue is to be involved in antigen contact.

Second, we have calculated the mean percentage burial over the accessible residues.

Click [here](#) for an image showing a composite combining site containing all CDR conformations coloured by contact propensity.

The table presents the chain name, residue number (N.B. This is **pre-1989 Chothia Numbering**), the number of contacts and the mean percent burial. The data are available by clicking [here](#).

An alternative simplified view is presented as a [list of CDR residues making contact](#) in each antibody with summary data for each CDR.

How to identify the CDRs by looking at a sequence

CDR-L1

Start - Approx residue 24

Residue before is **always** a Cys

Residue after is **always** a Trp. Typically TRP-TYR-GLN, but also, TRP-LEU-GLN, TRP-PHE-GLN, TRP-TYR-LEU

Length 10 to 17 residues

CDR-L2

Start - **always** 16 residues after the end of L1

Residues before generally ILE-TYR, but also, VAL-TYR, ILE-LYS, ILE-PHE

Length **always** 7 residues (except 7FAB which has a deletion in this region)

CDR-L3

Start - **always** 33 residues after end of L2 (except 7FAB which has the deletion at the end of CDR-L2)
Residue before is **always** Cys
Residues after **always** PHE-GLY-XXX-GLY
Length 7 to 11 residues

CDR-H1

Start - Approx residue 26 (**always** 4 after a CYS) [Chothia / AbM definition] Kabat definition starts 5 residues later
Residues before **always** CYS-XXX-XXX-XXX
Residues after **always** a TRP. Typically TRP-VAL, but also, TRP-ILE, TRP-ALA
Length 10 to 12 residues (AbM definition) Chothia definition excludes the last 4 residues

CDR-H2

Start - **always** 15 residues after the end of Kabat / AbM definition) of CDR-H1
Residues before typically LEU-GLU-TRP-ILE-GLY, but a number of variations
Residues after LYS/ARG-LEU/ILE/VAL/PHE/THR/ALA-THR/SER/ILE/ALA
Length Kabat definition 16 to 19 residues (AbM definition ends 7 residues earlier)

CDR-H3

Start - **always** 33 residues after end of CDR-H2 (**always** 2 after a CYS)
Residues before **always** CYS-XXX-XXX (typically CYS-ALA-ARG)
Residues after **always** TRP-GLY-XXX-GLY
Length 3 to 25(!) residues

Kabat Database and its applications: 30 years after the first variability plot

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ABSTRACT

The Kabat Database was initially started in 1970 to determine the combining site of antibodies based on the available amino acid sequences at that time. Bence Jones proteins, mostly from human, were aligned, using the now-known Kabat numbering system, and a quantitative measure, variability, was calculated for every position. Three peaks, at positions 24-34, 50-56 and 89-97, were identified and proposed to form the complementarity determining regions (CDR) of light chains. Subsequently, antibody heavy chain amino acid sequences were also aligned using a different numbering system, since the locations of their CDRs (31-35B, 50-65 and 95-102) are different from those of the light chains. CDRL1 starts right after the first invariant Cys 23 of light chains, while CDRH1 is eight amino acid residues away from the first invariant Cys 22 of heavy chains. During the past 30 years, the Kabat database has grown to include nucleotide sequences, sequences of T cell receptors for antigens (TCR), major histocompatibility complex (MHC) class I and II molecules and other proteins of immunological interest. It has been used extensively by immunologists to derive useful structural and functional information from the primary sequences of these proteins. An overall view of the Kabat Database and its various applications are summarized here. The Kabat Database is freely available at <http://immuno.bme.nwu.edu>

INTRODUCTION

The purpose of maintaining the Kabat Database of aligned sequences of proteins of immunological interest, in our opinion, is to provide useful correlations between structure and function for this special group of proteins from their nucleotide and amino acid sequences to their tertiary structures (1). These sequences are thus aligned with the ultimate aim of understanding how these proteins are folded and how they can perform their biological functions. We include only coding region sequences that have been published. In some cases, only the amino acid sequences were published, while the corresponding nucleotide sequences were deposited in GenBank. All stored

sequences were then printed out and checked visually against available published sequences. We routinely survey for possible new sequences in journals in our libraries, Medline entries, cross-references from other papers, and author notification; however, we may still miss some sequences. GenBank, on the other hand, contains a substantial number of unpublished sequences. If there are doubts about these sequences or their annotations, please refer to the original papers. The Kabat numbering systems (see the Introduction of 2) for antibody light and heavy chains, for TCR alpha and beta chains, etc., go hand-in-hand with variability calculations. The locations of the CDRs are the theoretically derived positions which can be verified experimentally. Indeed, from the first antigen-antibody Fab complex (3) to the complexes of TCR, processed peptide and MHC class I molecule (4,5), it has been realized that alignment of amino acid sequences and variability calculations can be of utmost importance in understanding how these important macromolecules function biologically. Due to the rapid development of genetic and protein engineering methods, mouse and rat antibodies have been humanized to treat human cancers, viral infections, etc (6). CDRs of selected rodent antibodies are cut out and glued onto human antibody frameworks to minimize rejection by human patients.

Our predicted CDRs are slightly different from Chothia's. A careful comparison can be found from a hyperlink on our website to 'Andrew's Antibody Page' (<http://www.biochem.ucl.ac.uk/~martin/abs/index.html>).

Massive amounts of sequence data are being continuously published in the scientific literature. It is imperative to collect and properly align the sequences so that they can be used by as many researchers in this field as possible. We have previously published five editions of these sequences (see the Introduction of 2). In 1991, the fifth edition (2) consisted of three volumes. Currently, the database is more than five times as large. As of September 29, 1999, the Kabat database contained 1 599 375 and 2 517 756 nt for antibody light and heavy chain variable regions, respectively, as compared to 272 244 and 418 962 nt in 1991. Total numbers of entries, amino acids and bases of other categories of sequences can be obtained by using the 'Current Counts' hyperlink on our website. The collection is available on our website (<http://www.immuno.bme.nwu.edu>) which is free due to the generous support by various research grants from NIH since 1970.

Finally, numerous scientific papers have cited our database, quoting our fourth edition (7), fifth edition (2), or one of our more recent papers (8). On our part, we have been analyzing

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the Kabat Database during the past few years with reference to the total numbers of antibody and TCR V-genes, possible evolutionary selection processes, importance of antibody CDRH3s as related to their fine specificities, etc.

KABAT DATABASE

The Kabat Database may be accessed for searching, sequence retrieval and analysis by a few different methods: electronic mail, WWW and ftp. The electronic mail interface has been available since 1993, the WWW interface since 1995 and various formats of the database in electronic format for nearly a decade (8). Our data formats, searching tools, output formats and database structures have gradually been adopted by other immunological databases and interfaces.

Electronic mail interface

An electronic mail interface (seqhunt2@immuno.bme.nwu.edu) provides a non-interactive method for searching and sequence retrieval (9). Sending mail to the server address with the single word 'help' (no quotes) in the message body returns instructions for using the server.

All sequences classes are searchable and returnable. The query format allows making AND/OR/NOT constructed restrictions on the database and amino acid and nucleotide sequence pattern matching with allowable differences. Requests are processed as they are received and depending on the network traffic, take ~1–2 min to be searched and returned to the sender. The returned format is a fixed-line length record of 80 or fewer characters per line for ease in visual inspection and processing by user-written scripts or programs. The characters are plain text.

The query format for the sent request consists of two parts. The first part contains directives for the server to follow while the second part contains specifications of the search. Specification of the extent of data returned, the number of documents to return, starting document and whether plain ASCII text or PostScript should be used in the return format may be entered. Further, one can direct the server to return a distribution, the variability or unaligned raw data for the search specified.

The second part of the query contains the search restrictions on the database. Words separated by AND and OR may be used, as well as searching functions, like nucleotide/amino acid pattern matching and positional restriction matching.

There are basically three steps in translating and performing a search on the Kabat Database: generate the question or query, translate it into a format the server can recognize and decide on the output options desired of the returned matches. For example, if matches of mouse kappa light chains of anti-phosphorylcholine antibodies are desired, the query and restriction on the database would be:

Begin

@mouse and kappa and phosphorylcholine

The '@' before mouse tells the server that matches of the species mouse are desired, rather than searching through the entire database record for instances of the word 'mouse'. More complicated restrictions can be generated using parentheses for grouping and the minus sign '-' for NOT. Finding all rat and rabbit sequences which are not kappa light chains, and returning them as amino acid sequences in PostScript format would be constructed as:

PSAA

Begin

(rat and rabbit) and -kappa

Pattern matching is interpreted as the second part of an AND statement, such that finding all rat and rabbit sequences which are not kappa and contain the nucleotide pattern cagtacgtcag with three allowable mismatches, would be sent as:

Begin

(rat and rabbit) and -kappa [implicit AND]

#NM 3

cagtacgtcag

More examples of searching and output options may be found in the 'help' file returned from the server.

WWW interface

The WWW interface (8) to the Kabat Database: <http://immuno.bme.nwu.edu> contains searching and analysis tools as well as links to database download sites and other interesting databases. Most of the features found in the electronic mail interface are found in the WWW interface, as well as other tools. The WWW interface is more interactive than the Email and returns results faster, depending on the network traffic.

Searching and analysis tools

SeqhuntII. This grouping of programs allows searches through the annotations and sequence pattern matching of the amino acid and nucleotide sequence data with allowable mismatches. Like the Email server, restrictions on the database may be formulated as AND/OR/NOT constructs. Output extent, output format, maximum documents and starting document may be specified. Browsing of the output results in HTML format allows the user to view the database entries in an easy-to-read format. ASCII text may be selected as output for use in user-generated scripts and programs. PostScript generation allows for printing on a PostScript supporting printer. Sequence matching is returned aligned with the target sequence with nucleotide or amino acid differences from the database sequence displayed in a case change. Since the database contains only coding regions of genes and proteins, the query sequence should be a portion of the coding region being sought.

Variability. Variability and amino acid distributions of sequence groups may be generated for restrictions on the database. The variability plots are in PostScript format and may either be viewed on the screen with an appropriate PostScript viewer (e.g. GNU ghostscript or ghostview) or printed to a postscript-supporting printer. Plots for human and mouse TCR gamma and delta chain variable regions are shown in Figure 1. Scaling of the variability plots may be done allowing comparison of variability plots for different groupings of sequences. Distributions of the amino acids per position may be returned also, including the calculated variability for each position.

Sequence alignment. Alignment of user-entered coding regions of immunoglobulin light chains according to the Kabat numbering system can be performed. Because of the relatively few alignment options available for light chains, most sequences can be aligned. One can start with around 10 amino acid residues or 30 nt. There is no lower limit on the length of sequence to be matched. In some cases though, visual inspection and alignment is necessary, as is for heavy chain alignment,

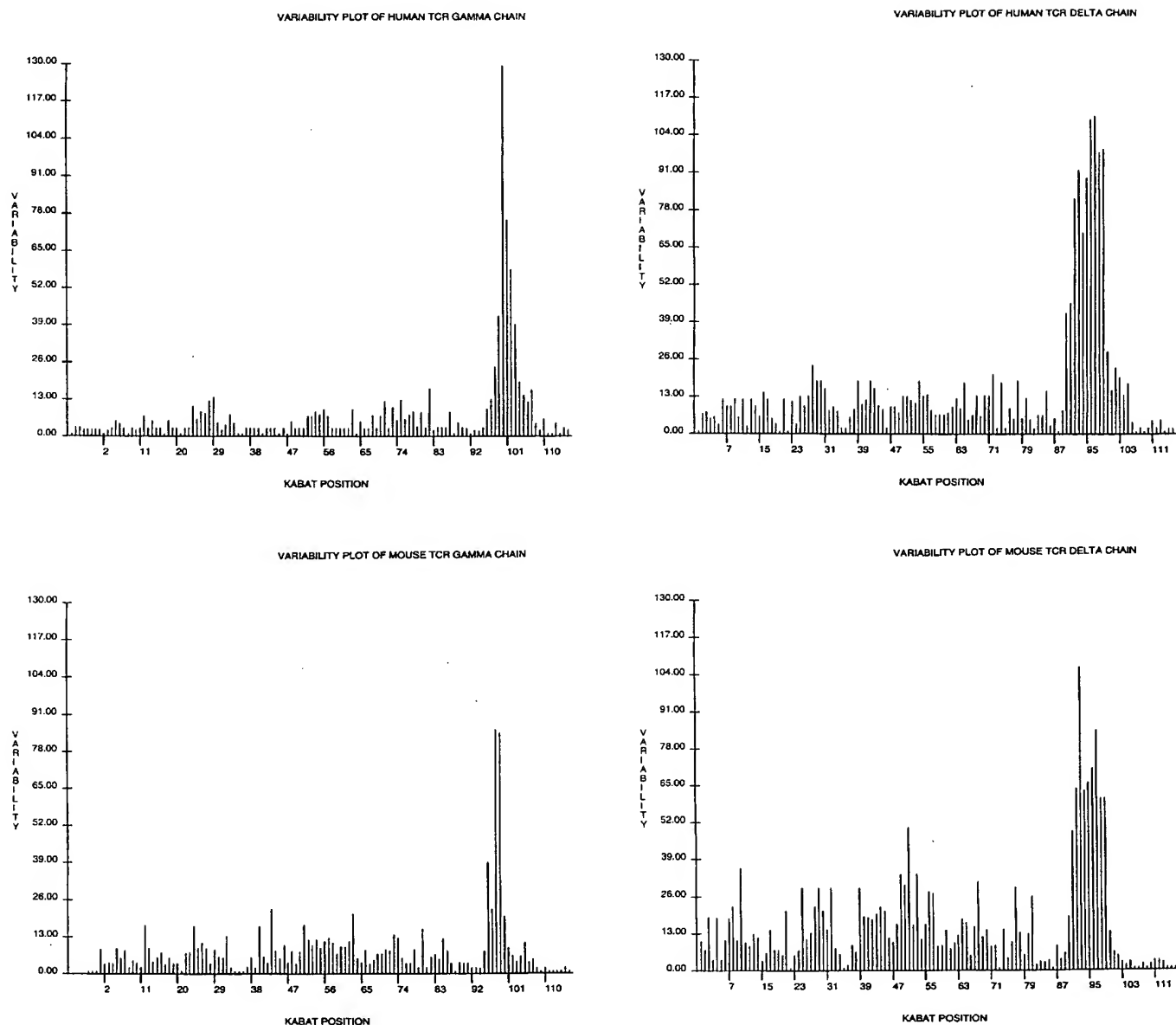


Figure 1. Variability plots for human and mouse TCR gamma and delta chain variable regions, using 377 human gamma, 1260 human delta, 297 mouse gamma and 461 mouse delta partial and complete sequences.

especially within the CDRH3 region, if additional codons or residues are inserted and denoted by '#'. If a suitable alignment counterpart from the database is not found for the target sequence, the user can contact us.

FTP. Various formats of the database are available for download from NCBI's repository under the directory 'kabat'. Currently active formats include a FASTA-like raw sequence format and the database's fixed length format of 80 or fewer

characters per line and vertical alignment. Four main variations on the fixed length format exist to properly visually display single translations, pseudogene translations, J-minigenes and D-minigenes. Other immunological databases have adopted similar formats as exemplified by the three letter code amino acid translation followed by single letter code. A 'dump' version of the database is periodically updated which contains unlimited line length records more suitable for mass processing on unix-based systems.

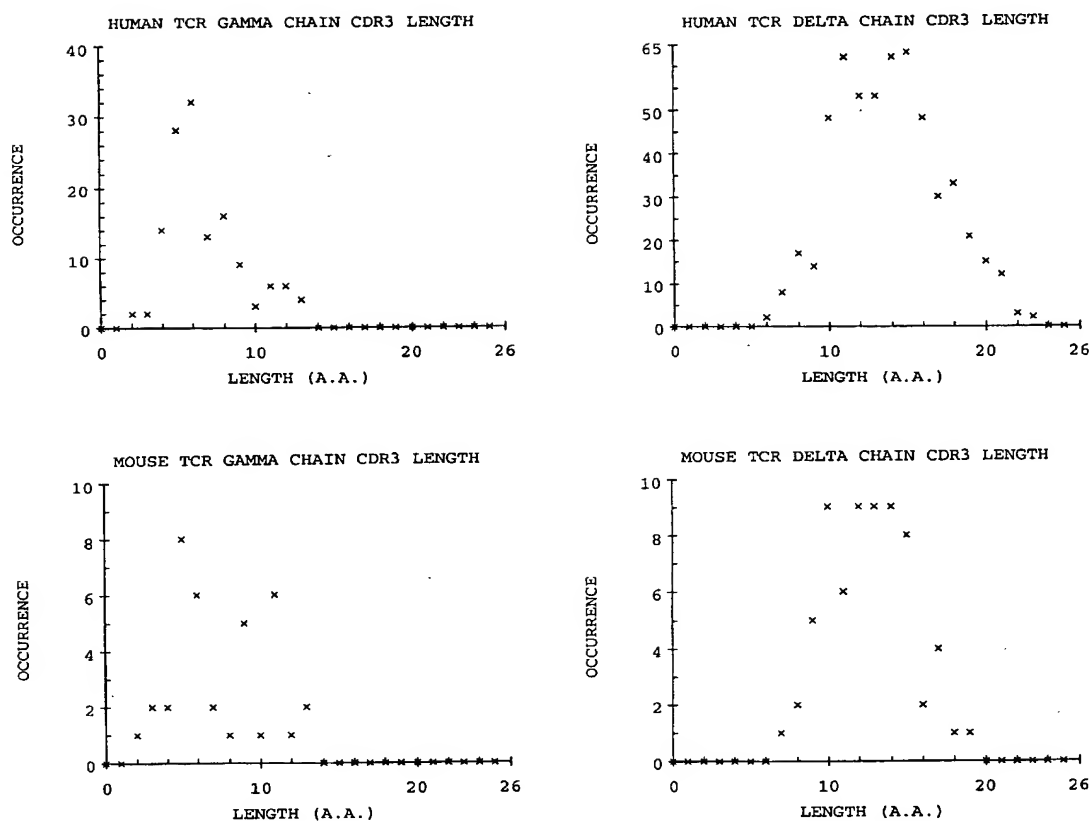


Figure 2. Length distributions of CDR3s of human and mouse TCR gamma and delta chains, based on 135 human gamma, 546 human delta, 37 mouse gamma and 66 mouse delta complete CDR3 sequences.

OTHER APPLICATIONS

As mentioned before, the Kabat Database was initially constructed for the purpose of identifying the antibody combining site (1). Starting from aligned amino acid sequences and using variability calculations, we have identified CDRs of antibody light and heavy chains, as well as those of TCRs. Such calculations can also provide useful predictions for MHC class I and II sequences (8), and to other aligned proteins sequences, e.g. HIV gp120, gp41, etc.

The importance of CDRH3 to confer fine specificity to antibodies was realized a few years ago (10). Furthermore, the unique CDRH3 nucleotide sequences have recently been used as a sensitive diagnostic test to detect residue B cell malignancies in cancer patients. Thus, many of these sequences have been determined. But most of them have been excluded from GenBank due to their relative short lengths. We have been meticulously collecting them, and realized the importance of their length distributions in antibodies of various specificities (11), and possible differences between CDRH3s of human and mouse (12). In the case of rabbit, the CDRH3s have less length variation than human and mouse. This may be compensated by the length variations of the CDRL3s (13).

The length variations of TCR alpha and beta chain CDR3s are very restricted (14). On the other hand, TCR gamma and delta chain CDR3s have more length variation, close to those of antibody heavy chains (Fig. 2). Whether they bind antigens directly is unclear.

During recent years, various research groups have decided to sequence the entire coding region of different antibody and TCR V-genes, in order to have an idea of their total numbers. On the other hand, we have discovered that pair-wise comparisons of V-gene nucleotide sequences in the Kabat Database provide very accurate estimations of their total numbers (15,16). In addition, such comparisons seem to suggest that antibody and TCR V-genes have evolved under different selective pressures (17). In the case of MHC class I sequences, comparison of their aligned sequences has elucidated a new mechanism of generating novel MHC class I molecules by random assortment of their $\alpha 1$ and $\alpha 2$ gene segments (18).

DISCUSSION

The Kabat Database has been around for 30 years. It has provided the immunology community a useful service, since it

not only is a sequence database but also incorporates vital aspects of the biology of the immune system. Various analytical methods have been developed to study the structure and function relations of proteins of immunological interest.

Electronic addresses:

<http://immuno.bme.nwu.edu>

seqhunt2@immuno.bme.nwu.edu

Citing the Kabat Database:

Authors using this database may cite this paper together with the electronic addresses.

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